

# Efficient purification of high quality viral nucleic acids from a wide range of sample materials using GENTi™ Advanced Viral DNA/RNA Extraction Kit

## Experimental Conditions

### Materials Required

- ♦ GENTi™ 32 Advanced Automatic Extraction System (GTI032A)
- ♦ GENTi™ Advanced Viral DNA/RNA Extraction Kit (902-048A/902-096A)
- ♦ 5 ml conical tube
- ♦ Pipette & sterile pipette tips
- ♦ Suitable protector (e.g., lab coat, disposable gloves, goggles, etc.)

### Sample Information

- ♦ Sample type: cultured virus and bacteria
  - Infectious bronchitis virus (IBV,  $10^{3.5}$  EID<sub>50</sub>)
  - Rabies virus (RV,  $10^{3.5}$  LD<sub>50</sub>)
  - Japanese encephalitis virus (JEV,  $10^{5.0}$  TCD<sub>50</sub>)
  - *Mycoplasma gallisepticum* (MG,  $1 \times 10^5$  CCU)
- ♦ Extraction conditions
  - Sample amount: 200 µl
  - Elution volume: 100 µl
  - Extraction protocol: Viral\_Normal (operation time : 29' 35")

### Sample Preparation

1. Mix the all cultured virus and bacteria medium to 5 ml conical tube and extract the 200 µl sample from the mixture.
2. One sample is according to GENTi™ Advanced Viral DNA/RNA Extraction Kit protocol, the other sample is according to manual method of viral DNA/RNA extraction kit each from two different suppliers for comparison.

## Protocol

### GENTi™ Advanced Viral DNA/RNA Extraction Kit Protocol

\* For more details and methods, please refer to [the handbook of GENTi™ Advanced Viral DNA/RNA Extraction Kit](#).

1. Add 7 µl of dissolved Carrier RNA (1 µg/µl) to 1st/7th well of plate type cartridge.
2. Add 200 µl of samples to 1st/7th well.
3. Load the plate type cartridge on the tray of GENTi™ 32 Advanced Automatic Extraction System.
4. Insert the magnetic rod cover to the end to strip bracket.
5. Select the correct extraction protocol and operate the extraction system.

## Result

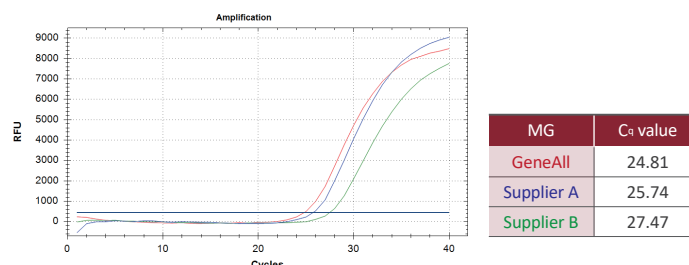


Figure 1. Comparison of C<sub>t</sub> value of DNA template extracted from the *Mycoplasma gallisepticum* (MG).

The DNA template was extracted from the *Mycoplasma gallisepticum* (MG) using GENTi™ Advanced Viral DNA/RNA Extraction Kit on GENTi™ 32 Advanced Automatic Extraction System (red line) and viral DNA/RNA extraction kit (manual method) each from two different suppliers (blue and green line). Eluted DNA template was analyzed with a TaqMan-based real-time PCR assay using CFX-96.

- Red line: GENTi™ Advanced Viral DNA/RNA Extraction Kit
- Blue line: supplier A manual extraction kit
- Green line: supplier B manual extraction kit

- Real-time PCR system: CFX-96™ System (1855201)
- qPCR kit: Probe qPCR Mix (RR391A)
- Target gene: none specific

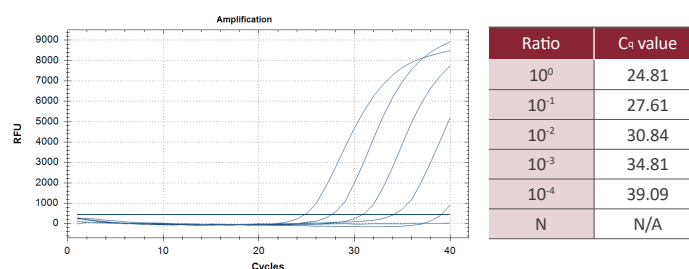
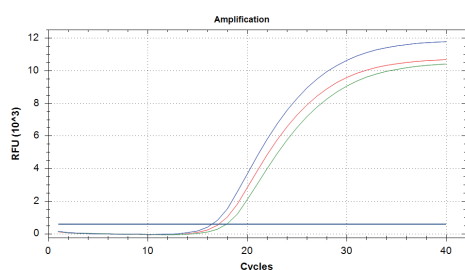


Figure 2. Analysis of extraction sensitivity on serial dilutions of *Mycoplasma gallisepticum* (MG).

The DNA template was extracted from a 10-fold serial dilution of *Mycoplasma gallisepticum* (MG) using GENTi™ Advanced Viral DNA/RNA Extraction Kit on GENTi™ 32 Advanced Automatic Extraction System. All eluates were analyzed with a TaqMan-based real-time PCR assay using CFX-96.

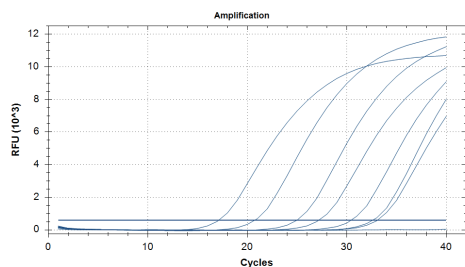
- N: negative control (nuclease-free water)
- Real-time PCR system: CFX-96™ System (1855201)
- qPCR kit: Probe qPCR Mix (RR391A)
- Target gene: none specific

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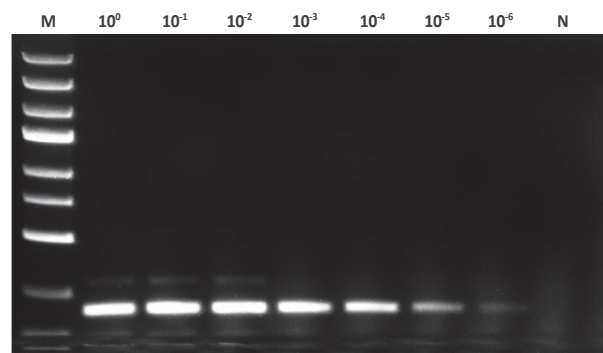
MG	C <sub>A</sub> value
GeneAll	17.14
Supplier A	16.45
Supplier B	17.92

**Figure 3. Comparison of C<sub>A</sub> value of RNA template extracted from the Infectious bronchitis virus (IBV).** The RNA template was extracted from the Infectious bronchitis virus (IBV) using GENTi™ Advanced Viral DNA/RNA Extraction Kit on GENTi™ 32 Advanced Automatic Extraction System (red line) and viral DNA/RNA extraction kit (manual method) each from two different suppliers (blue and green line). Eluted RNA template was synthesized to cDNA with reverse transcription; and then analyzed with TaqMan-based one-step RT-qPCR assay using CFX-96.  
 Red line: GENTi™ Advanced Viral DNA/RNA Extraction Kit  
 Blue line: supplier A manual extraction kit  
 Green line: supplier B manual extraction kit  
 • Real-time PCR system: CFX-96™ System (1855201)  
 • RT-qPCR kit: HyperScript™ One-Step RT-PCR Master Mix, 0.5 ml x 2 (602-125)  
 • Target gene: none specific

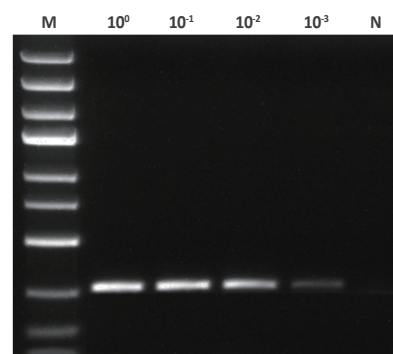


Ratio	C <sub>A</sub> value
10 <sup>0</sup>	17.14
10 <sup>-1</sup>	20.76
10 <sup>-2</sup>	25.01
10 <sup>-3</sup>	27.19
10 <sup>-4</sup>	30.44
10 <sup>-5</sup>	32.61
10 <sup>-6</sup>	33.09
N	N/A

**Figure 4. Analysis of extraction sensitivity on serial dilutions of Infectious bronchitis virus (IBV).** The RNA template was extracted from a 10-fold serial dilution of Infectious bronchitis virus (IBV) using GENTi™ Advanced Viral DNA/RNA Extraction Kit on GENTi™ 32 Advanced Automatic Extraction System. All elutes were analyzed with TaqMan-based one-step RT-qPCR assay using CFX-96.  
 • N: negative control (nuclease-free water)  
 • Real-time PCR system: CFX-96™ System (1855201)  
 • RT-qPCR kit: HyperScript™ One-Step RT-PCR Master Mix, 0.5 ml x 2 (602-125)  
 • Target gene: none specific



**Figure 5. Analysis of extraction sensitivity on dilutions of Rabies virus (RV).** The RNA template was extracted from a 10-fold serial dilution of known positive Rabies virus (RV) samples using GENTi™ Advanced Viral DNA/RNA Extraction Kit on GENTi™ 32 Advanced Automatic Extraction System. All eluates were analyzed using conventional reverse transcription PCR (RT-PCR) assay.  
 • N: negative control (nuclease-free water)  
 • Target gene (PCR product size): Jecom (100 bp)  
 • PCR system: MultiGene™ Optimax thermal cycler (TC9610)  
 • RT-PCR kit: HyperScript™ One-Step RT-PCR Master Mix, 0.5 ml x 2 (602-125)  
 • Electrophoresis conditions: 1.2% agarose, 110 V, 30 min, 10 µl



**Figure 6. Analysis of extraction sensitivity on dilutions of Japanese encephalitis virus (JEV).** The RNA template was extracted from a 10-fold serial dilution of known positive Japanese encephalitis virus (JEV) samples using GENTi™ Advanced Viral DNA/RNA Extraction Kit on GENTi™ 32 Advanced Automatic Extraction System. All eluates were analyzed using conventional reverse transcription PCR (RT-PCR) assay.  
 • N: negative control (nuclease-free water)  
 • Target gene (PCR product size): omRABV (192 bp)  
 • PCR system: MultiGene™ Optimax thermal cycler (TC9610)  
 • RT-PCR kit: HyperScript™ One-Step RT-PCR Master Mix, 0.5 ml x 2 (602-125)  
 • Electrophoresis conditions: 1.2% agarose, 110 V, 30 min, 10 µl